

# Endophytic Fungi as latent Pathogens in *Eichhornia Crassipes* (Mart.) Solms

Soumya P. R<sup>1</sup>, Rukshana Begum S<sup>2</sup> and Tamil Selvi K. S<sup>3</sup>

<sup>1</sup>Research Scholar, Department of Botany, PSGR Krishnammal College for Women, Coimbatore, Tamil Nadu – 641004, India.

<sup>2</sup>Research Scholar, Department of Botany, PSGR Krishnammal College for Women, Coimbatore, Tamil Nadu – 641004, India.

<sup>3</sup>Assistant Professor, Department of Botany, PSGR Krishnammal College for Women, Coimbatore, Tamil Nadu – 641004, India.

## Abstract

*Eichhornia crassipes* (water hyacinth) are one of the most invasive aquatic weed. Endophytic fungi and pathogenic fungi were isolated from *E. crassipes*. Fungi isolated as endophytes were tested to find if they are capable of causing disease symptoms on healthy leaves of *E. crassipes*. Endophytes, *Aspergillus flavus*, *A. fumigatus*, *A. glaucus*, *Curvularia lunata*, *Eurotium repens*, *Oidium* sp., *Chaetomium globosum* and sterile mycelia showed disease symptoms on healthy leaves. The results confirmed that endophytic fungi may act latent pathogen in their host plant under certain stress conditions.

**Key words:** *Eichhornia crassipes*, endophytic fungi, pathogenic fungi, latent pathogens.

## 1. Introduction

In India, many rivers, irrigation canals, lakes both natural and man - made, are choked by the explosive growth of aquatic weeds, resulting in enormous direct losses. Besides different types of algae, the most important representatives of aquatic weeds in India are: *Eichhornia crassipes* (free floating), *Nymphaea stellata* (rooted floating), *Nelumbo nucifera* (rooted floating), *Hydrilla verticillata* (root submerged), *Typha angusta* (emergent), etc. (Varshney *et al.*, 2008).

*Eichhornia crassipes* (Mart.) Solms, commonly known as water hyacinth is considered as the most predominant, troublesome aquatic weed (Pathak and Kannan, 2015) and listed as one of the most productive plant on the earth (Kayathri *et al.*, 2015). The plant is otherwise called as “Blue Devil” or “Bengal terror” in India (Bhattacharya *et al.*, 2015). It has been proven to be a persistent and

expensive weed problem all over the world (Sotolu, 2012).

Plants serve as reservoirs of large numbers of microorganisms known as endophytes (Bacon and White, 2000); that colonize the living tissues of plants without causing any negative effect on the host plant (Hirsch and Broun, 1992). They are the best example of positive plant - microbe interaction and association of different plant species with fungi and bacteria (Hallmann, 1997). The term “endophyte” was coined by German phytopathologist Heinrich Anton De Bary in 1884, and is used to define fungi or bacteria living inside plant tissues without showing any disease symptoms in the host plant (Wilson, 1995, Azevedo, 1998). The word endophyte has come from two Greek words, “endon” means within and “phyton” means plant. They play a vital role and constitute an important component of plant micro-ecosystems (Tan and Zhou, 2001; Rodriguez *et al.*, 2009).

Endophytic fungi play a role in plant protection from pests and insects, but at the same time they can be pathogenic (Weber, 1981; Malinowski and Belesky, 2006). Under certain conditions endophytes may become parasitic, causing symptomatic infection in their host (Brown *et al.*, 1998). The disease symptoms of host plant can be caused by endophytes under stress conditions (Clay and Schardl, 2002; Schulz and Boyle, 2005). It can be regarded as an unbalanced status of a symbiosis when the host is stressed and physiological or ecological conditions favors virulence (Muller *et al.*, 2005; Schulz and Boyle, 2005; Kogel *et al.*, 2006). Endophytes of certain plant could be a pathogen of other plants, depending on the balance between pathogenicity and endophytism of the microorganism in the

different hosts (Saikkonen *et al.*, 2004). Among all aquatic weeds, fungal pathogens associated with water hyacinth have been extensively studied (Tessmann *et al.*, 2001).

Endophytes may turn into a pathogen in response to some environmental cue (Hendry *et al.*, 2002); such a shift in the nature of the endophyte also result in a change in its metabolite profile. The environmental conditions which affect host plant growth, influence the number and variety of endophytic populations, and also affect metabolites production in endophytes. The purpose of this study was to identify endophytic fungal taxa that might exist as latent pathogens in *E. crassipes*.

## 2. Materials and Methods

### 2.1 Isolation of endophytic fungi

Endophytic fungi were isolated from leaves, petioles and roots of healthy and asymptomatic tissues of *E. crassipes* from Singanallur and Ukkadam wetlands, Coimbatore. The collected plant samples were surface sterilized, cut into 1 cm long and 3-4 mm broad pieces and aseptically transferred to PDA medium. The plates were incubated at 25°C for 7 days and observed regularly for fungal growth. The mycelial outgrowth from each segment was separated and subcultured for further study.

### 2.2 Isolation of pathogenic fungi

Pathogenic fungi were isolated from diseased and infected leaves of *E. crassipes* from Singanallur Lake. The collected plant samples were surface sterilized, cut into 1×1 cm pieces and aseptically transferred to PDA medium. The plates

were incubated at 27°C for 7 days and observed regularly for fungal growth.

### 2.3 Pathogenicity tests of endophytic fungi

The isolated endophytes were subjected to pathogenicity test to check if they act as latent pathogens on leaves of *E. crassipes*. Fungal isolates were grown on potato dextrose agar medium for one week. Healthy leaves of *E. crassipes* were washed under running tap water, surface sterilized, placed in sterile petriplates layered with sterilized moist tissue paper. Endophytic fungi were reinoculated into a healthy leaves by wounding technique and incubated for 1week under room temperature.

## 3. Results

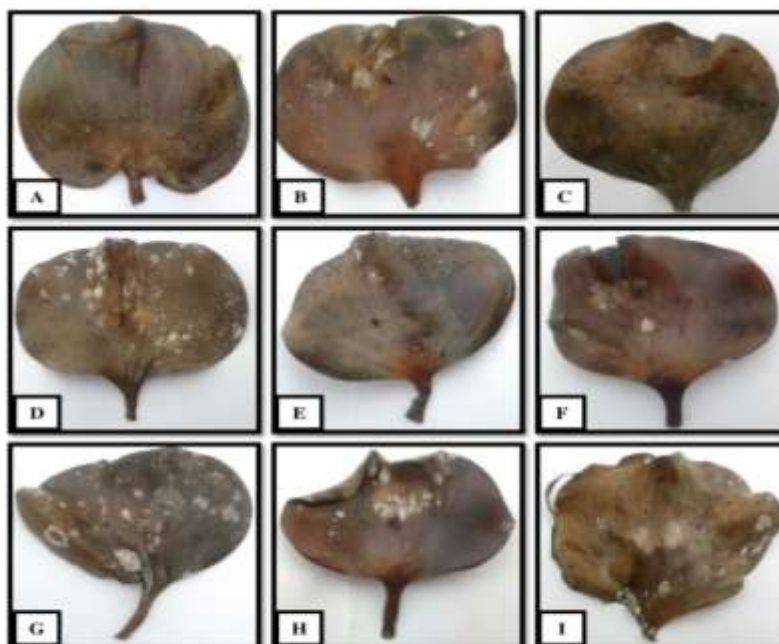
Thirteen endophytic fungi were isolated from leaves, petioles and roots of *E. crassipes* and 5 pathogenic fungi were isolated from infected leaves of *E. crassipes* (Table 1).

*Aspegillus flavus* and *Curvularia lunata* were isolated in both healthy and infected leaves of *E. crassipes*.

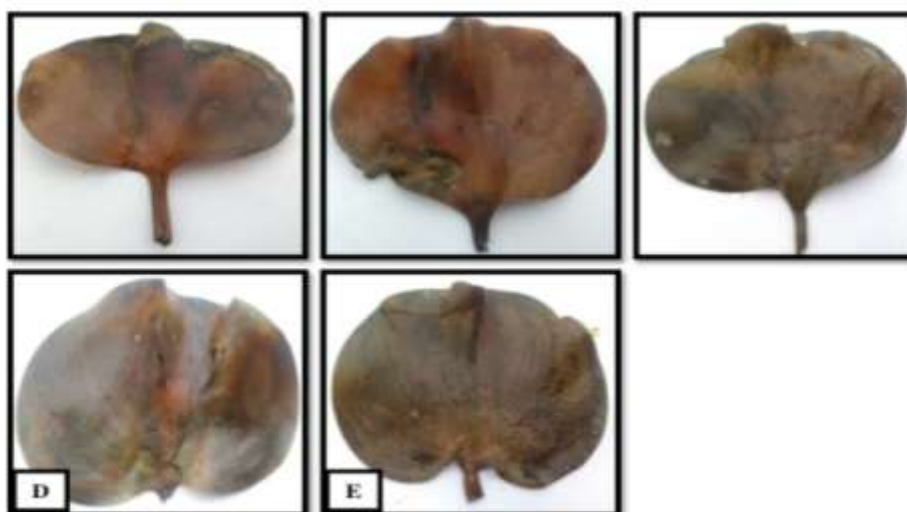
Thirteen endophytic fungi were tested for their pathogenicity on leaves of *E. crassipes*. The fungi were inoculated into the leaves by artificial wounding method. After one week, *Aspegillus flavus*, *A. fumigatus*, *A. glaucus*, *Curvularia lunata*, *Eurotium repens*, *Oidium* sp., *Chaetomium globosum* and sterile mycelia showed disease symptoms on leaves of *E. crassipes*. The other five species and control did not produce any disease symptoms (Figure 1 & 2).

**Table 1: Endophytic and pathogenic fungi isolated from *E. crassipes***

Endophytic fungi	Pathogenic fungi
<i>Acremonium bolchii</i>	<i>Acremonium strictum</i>
<i>Acremonium strictum</i>	<i>Aspegillus flavus</i>
<i>Aspegillus flavus</i>	<i>Curvularia lunata</i>
<i>Aspegillus fumigatus</i>	<i>Penicillium digitatum</i>
<i>Aspegillus glaucus</i>	<i>Penicillium restrictum</i>
<i>Aspegillus japonicus</i>	
<i>Chaetomium globosum</i>	
<i>Curvularia lunata</i>	
<i>Eurotium repens</i>	
<i>Oidium</i> sp.	
<i>Penicillium digitatum</i>	
<i>Penicillium restrictum</i>	
Sterile mycelia	



**Figure-1.** Pathogenicity test of endophytic fungi: Typical disease symptoms on healthy leaves of *E. crassipes* when endophytic isolates were inoculated by wounding method.  
**A.** Control **B** *Aspergillus flavus* **C.** *Aspergillus fumigatus* **D.** *Aspergillus glaucus*  
**E.** *Chaetomium globosum* **F.** *Curvularia lunata* **G.** *Eurotium repens* **H.** *Oidium* sp.  
**I.** Sterile mycelia.



**Figure 2:-** Pathogenicity test of endophytic fungi: Without showing disease symptoms on healthy leaves of *E. crassipes* when endophytic isolates were inoculated by wounding method.  
**A.** *Acermonium bolchii* **B** *Acremonium strictum* **C.** *Aspergillus japonicus*  
**D.** *Penicillium digitatum* **E.** *Penicillium restrictum*

#### 4. Discussion

*Eichhornia crassipes* is considered one of the world's worst aquatic plants (Toft, 2000). Out of the 160 aquatic weeds, *Eichhornia crassipes* is of primary concern in India (Sushilkumar, 2011). The endophytic fungi spend the whole or part of their life cycle in healthy living tissues of host plant by colonizing inter or intra- cellularly without

causing any apparent symptoms of diseases (Azevedo, 1998). Evidences that an endophytic fungi showed disease symptoms in host plants under certain conditions has been discussed by several authors (Clay and Schardl, 2002; Schulz and Boyle, 2005). Thus the purpose of the study was establish if any fungi isolated as endophytes

from *E. crassipes* have the ability to be latent pathogens.

Thirteen endophytic fungi and 5 pathogenic fungi were isolated. Genera that include common endophytes and isolated as pathogens from *E. crassipes* were *Acremonium*, *Aspergillus*, *Curvularia* and *Penicillium*. *Curvularia lunata* was

isolated in both healthy and infected leaves of *E. crassipes*. Fungal pathogens associated with water hyacinth have been extensively studied (Tessmann *et al.*, 2001). *Curvularia lunata* is already reported as serious pathogen of water hyacinth (Pathak and Kannan, 2015) and number of Pathogens had previously recorded from *E. crassipes* (Table 2).

**Table 2: Fungi previously recorded as pathogens of *E. crassipes***

Fungi	Reference
<i>Alternaria eichhorniae</i> , <i>Myrothecium roridum</i> and Visarathanonth, 1975	Rakvidhasatra and <i>Rhizoctonia solani</i>
<i>Myrothecium</i> spp., <i>Rhizoctonia</i> spp. and <i>Pestalotia</i> sp.	Syed, <i>et al.</i> , 1978
<i>Helminthosporium</i> , and <i>Chaetomella</i>	Caunter, 1982
<i>Alternaria alternate</i> , * <i>Aspergillus niger</i> , ** <i>Curvularia lunata</i> , <i>Fusarium oxysporum</i> , * <i>Penicillium chrysogenum</i> , <i>Pythium</i> sp., <i>Trichoderma viride</i> , <i>Trichothecium</i> sp.	Pathak and Kannan, 2011

Note: \*Genus isolated as endophytes \*\*Species isolated as endophytes

Some pathogens have a latent phase within the host tissue and some saprobes can also be facultative parasites (Millar, 1980; Andrews *et al.*, 1985). Endophytes and pathogens both possess many of the same virulence factors. Therefore, they can produce similar symptoms (Schulz *et al.*, 2002; Seier and Romero, 1996, 1997). However, fungal endophytes are closely related to plant pathogens, the fungi can either biotrophic fungi or necrotrophic fungi in their host plant (Delaye *et al.*, 2013).

Endophytes have isolated by plant pathologists and described as weak pathogens under physiological stress conditions (Kulik, 1984). Brown *et al* (1998) suggested that endophytes are also thought to be latent pathogens and were slightly pathogenic to the host plant (Padhi *et al.*, 2013). The endophytic fungi *Acremonium strictum*, *A. bolchii*, *A. flavus*, *A. fumigatus*, *A. glaucus*, *A. japonicus*, *Curvularia lunata*, *Eurotium repens*, *Chaetomium globosum*, *Oidium* sp., *P. digitatum*, *P. restrictum* and sterile mycelia were tested for their pathogenicity on leaves of *E. crassipes*. Among the thirteen endophytes, *A. flavus*, *A. fumigatus*, *A. glaucus*, *Curvularia lunata*, *Eurotium repens*, *Oidium* sp., *Chaetomium*

*globosum* and sterile mycelia showed disease symptoms on leaves. These results were similar to earlier works of Wright, 1998. Endophytic fungi act latent pathogens were previously reported in plants (Table 3).

A latent phase represents a specific condition where the fungus can either develop symptoms or cause change in their physiology of host plant (Romero *et al.*, 2001). Several researchers reported that endophytes may become latent pathogen due to changes in environmental conditions such as CO<sub>2</sub> accumulation or O<sub>2</sub> depletion (Lund and Wyatt, 1972). They have evolved directly from plant pathogenic fungi (Carroll, 1988; Isaac, 1992).

*Acremonium strictum*, *P. digitatum* and *P. restrictum* were did not produce any symptoms of disease in *E. crassipes*. Praveena and Naseema (2004) reported that, *Penicillium* sp. and *Aspergillus* sp. were non pathogenic. True endophytic fungal colonization never showed any visible disease symptoms in their host plant (Mostert *et al.*, 2000). Endophytic fungi are capable of living in host plants without causing any symptoms (Petrini *et al.*, 1992).



**Table 3: Previous study showed endophytic fungi act latent pathogens in plants**

Endophytic fungi	Name of plant	Disease
<i>Phomopsis citri</i>	<i>Citrus</i> spp.	Stem end rot (Wright, 1998)
<i>Fusicoccum aesculi</i>	<i>Citrus</i> spp.	Stem end rot (Wright, 1998)
<i>Lasiodiplodia theobromae</i>	<i>Citrus</i> spp.	Stem end rot (Wright, 1998)
<i>Phomopsis viticola</i>	<i>Vitis vinifera</i>	Leaf lesions (Mostert <i>et al.</i> , 2000)
<i>Deightonella torulosa</i>	<i>Musa acuminata</i>	Leaf spots (Photita <i>et al.</i> , 2004)

## 5. Conclusion

Endophytic fungi and pathogenic fungi were isolated from healthy and infected tissues of *E. crassipes* respectively. Fungi isolated as endophytes were tested for their pathogenicity on healthy leaves of *E. crassipes*. Thus present investigation revealed that endophytic fungi acted latent pathogens in healthy leaves of *E. crassipes*.

## Acknowledgement

We thankful to the Head of the Department of Botany PSGR Krishnammal College for Women, Coimbatore for providing facilities in the laboratory.

## References

- [1] Andrews JH, Hecht EP and Bashirian S, Association between the fungus *Acremonium curvulum* and Eurasian water milfoil, *Myriophyllum spicatum*. Canadian Journal of Botany, 60: 1216-1221, 1985.
- [2] Azevedo JL Microorganismos as endofiticos. In: Melo IS and Azevedo J. L., editors. Ecologia Microbiana, Brazil: Editora Embrapa, Jaguariuna, Sao Paulo. pp. 117-137, 1998.
- [3] Bacon CW and White JF. Microbial Endophytes. Marcel Dekker, New York. Pp.341-388, 2000.
- [4] Bhattacharya A, Haldar S and Chatterjee PK, Geographical distribution and physiology of water hyacinth (*Eichhornia crassipes*) – the invasive hydrophyte and a biomass for producing xylitol. International Journal of ChemTech Research, 7 (4): 849-1861, 2015.
- [5] Brown KB and Hyde KD, Guest DJ, Preliminary studies on endophytic fungal communities of *Musa acuminata* species complex in Hong Kong and Australia. Fungal Diversity, 1: 27-51, 1998.
- [6] Carroll GC, Fungal endophytes in stem and leaves: from latent pathogen to mutualistic symbionts. Ecology, 69:2-9, 1988.
- [7] Caunter IG, Potential for biological control of water hyacinth with indigenous fungal pathogens. Malaysian plant protection society, 489-492, 1982.
- [8] Clay K and Schardl CL, Evolutionary origins and ecological consequences of endophyte symbiosis with grasses. American Naturalist, 160: 99-127, 2002.
- [9] Delaye L, García-Guzmán G and Heil M, Endophytes versus biotrophic and necrotrophic pathogens -are fungal lifestyles evolutionarily stable traits? Fungal Diversity DOI 10.1007/s13225-013-0240-y. 2013.
- [10] Hallmann J, Quadt-Hallman A, Mahaffee, WF and Kloepper, JW, Bacterial endophytes in agriculture crops. Can J Microbiol 43:895-914, 1997.
- [11] Hendry SJ, Boddy L, Lonsdale D, Abiotic variables effect differential expression of latent infections in beech (*Fagus sylvatica*). New Phytologist, 155: 449-460, 2002.
- [12] Hirsch G and Broun U, Communities of parasitic microfungi. In: Winterhoff W, editor. Handbook of vegetation science. Kluwer Academic Publisher 225-250, 1992.
- [13] Isaac S, Fungal plant interactions. Chapman and Hall, London. 418, 1992.
- [14] Toft JD, Community Effects on the Non-Indigenous Aquatic Plant Water Hyacinth (*Eichhornia crassipes*) in the Sacramento/San Joaquin Delta, California. 1-86, 2000.
- [15] Kayathri B, Kanimozhi K and Panneerselvam A, Preliminary phytochemical analysis and In vitro

- investigation of antimicrobial activity of *Eichhornia crassipes* (Mart.) Solms. Against poultry pathogens. CIBTech Journal of Microbiology, 4 (1): 19-27, 2015.
- [16] Kogel KH, Franken P and Huckelhoven R, Endophyte or parasite – what decides? Current Opinion in Plant Biology, 9: 358–363, 2006.
- [17] Kulik MM, Symptomsless infection, persistence, and production of pycnidia in host and non-host plants by *Phomopsis batatae*, *Phomopsis phaseoli* and *Phomopsis sojiae* and the taxonomic implications. Mycologia, 76: 274-291, 1984.
- [18] Lund B M. and Wyatt GM, The effect of oxygen and carbon dioxide concentrations on bacterial soft rot of potatoes. I. King Edward potatoes inoculated with *Erwinia carotovora* var. *atroseptica*. Potato Res, 15:174-179, 1972.
- [19] Malinowski DP and Belesky DP, Ecological importance of *Neotyphodium* spp. Grass endophytes in agroecosystems. Grassland Science, 52(1): 23-28, 2006.
- [20] Millar CS, Infection processes on conifer needles. In: Microbial Ecology of the Phylloplane (ed. J.P. Blakeman). Academic Press: London, UK: 185-209, 1980.
- [21] Mostert L, Crous PW and Petrini O, Endophytic fungi associated with shoots and leaves of *Vitis vinifera*, with specific reference to the *Phomopsis viticola* complex. Sydowia, 52: 46- 58. 2000.
- [22] Muller CB and Krauss J, Symbiosis between grasses and asexual fungal endophytes. Current Opinion in Plant Biology, 8:450-456, 2005.
- [23] Padhi L, Mohanta YK and Panda SK, Endophytic fungi with great promises: A review. Journal of Advanced Pharmacy Education and Research, 3(3): 152-170, 2013.
- [24] Pathak, A. and Kannan C, Isolation and pathogenicity of some native fungal pathogens for the biological management of water hyacinth. Indian Journal of Weed Science, 43: (3&4)178-180, 2011.
- [25] Photita W, Lumyong S, Lumyong P, McKenzie, EHC and Hyde KD, Are some endophytes of *Musa acuminata* latent pathogens? Fungal Diversity 16: 131-140, 2004.
- [26] Petrini O, Sieber T, Toti, L and Vire O, Ecology, metabolite production and substrate utilization in endophytic fungi. Nat Toxins, 1:185–196, 1992.
- [27] Praveena R and Naseema A, Fungi occurring on water hyacinth [*Eichhornia crassipes* (Mart.) Solms] in Kerala. Journal of Tropical Agriculture, 42(1-2): 21-23, 2004.
- [28] Raknidhyasastra V and Visarathanonh, Isolation and identification of fungi pathogenic to waterhyacinth (*Eichhornia crassipes* [Mart.] Solms.). Kasetsart. J, 9:170-7, 1975.
- [29] Rodriguez RJ, White, JF, Arnold, AE and Redman RS, Fungal endophytes: Diversity and functional roles. New Phytologist, 182: 314-330, 2009.
- [30] Romero A, Carrión G and Rico-Gray V, Fungal latent pathogens and endophytes from leaves of *Parthenium hysterophorus* (Asteraceae). Fungal Diversity, 7: 81-87, 2001.
- [31] Saikkonen K, Wali, P, Helander and M Faeth SH, Evolution of endophyte-plant symbioses. Trends in Plant Science, 9: 275-280, 2004.
- [32] Schulz B and Boyle C, The endophytes continuum. Mycological Research, 109:1661-686, 2005.
- [33] Schulz B, Boyle C, Draeger S, Rommert AK and Krohn K, Endophytic fungi: a source of novel biologically active secondary metabolites. Mycol Res, 106:996-1004, 2002.
- [34] Seier MA and Romero, Control of *Parthenium* weed (*Parthenium hysterophorus*), July, 1995 - June 1996. Annual Report, IIBC, 1996.
- [35] Seier MA and Romero, Control of *Parthenium* weed (*Parthenium hysterophorus*), July, 1996 - June 1997. Annual Report, IIBC, 1997.
- [36] Sotolu AO, Management and Utilization of Weed: Water hyacinth (*Eichhornia crassipes*) for Improved Aquatic Resources. Journal of Fisheries and Aquatic Science, 1-8, 2012.
- [37] Sushilkumar, Aquatic weeds problems and management in India. Indian Journal of Weed Science, 43 (3&4): 118-13, 2011.
- [38] Syed RA, Setyawati O and Kasno, Biological control of some aquatic weeds (for BIOTROP). Trinidad, Commonwealth Institute of Biological Control, Report of work carried out during 1976. 187-189, 1978.
- [39] Tan RX and Zhou WX, Endophytes: a rich source of functional metabolites. Nat. Prod. Rep, 18: 488-459, 2001.

- [40] Tessmann DJ, Charudattan R, Kistler HC and Roskopf E A molecular characterization of *Cercospora* species pathogenic to waterhyacinth and emendation of *C. piaropi* Tharp. *Mycologia*, 93: 323-34, 2001.
- [41] Varshney JG, Shushilkumar and Mishra JS, Current Status of Aquatic and Their management in India. Sengupta, M. and Dalwani, R. (Editors.). *Proceedings of Taal* 2007: The 12<sup>th</sup> World Lake Conference: 1039-1045, 2008.
- [42] Weber J, A natural control of Dutch elm disease. *Nature London*, 292:449-451, 1981.
- [43] White JF Jr, The wide spread distribution of endophytes in the poaceae. *Plant disease*, 71: 340 – 342, 1987.
- [44] Wilson D, Endophyte: the evolution of a term and clarification of its use and definitions. *Oikos*, 73: 274–276, 1995.